



Prescription poop: The microbiome, dysbiosis and FMT

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Intestinal microbiota

Courtesy of Jan Suchodolski GI LAB Texas A&M





Highly diverse and balanced ecosystem

Bacteria most represented with100 x 10¹² microorganisms from >1000 species

Metabolic function

Beneficial metabolites

Short chain fatty acids (SCFA) – from dietary NFE/fiber

Faecalibacterium prausnitzii & others

Tryptophan + lipid metabolites (indole, kynurenine and kynurenic acid, sphingolipids) *Many bacterial species*

Secondary bile acids (SBA) – from primary bile acids

Clostridium (Peptacetobacter) hiranonis (dogs & cats)



Intestinal dysbiosis

Changes in composition and/or function of the intestinal microbiota



DeGruttola AK, et al. Inflamm Bowel Dis. 2016 May;22(5):1137-50

Disruptors of GI homeostasis

Poor quality and/or imbalanced diet*

Enteropathogens (CPV, FPV, FeCV)*

Antimicrobials*, NSAIDs, PPIs

Inflammation

especially early in life*

Suchodolski JS - In "Purina Institute - Canine and Feline Clinical Nutrition Handbook, 2023 edition"



Dysbiotic and reduced bacterial diversity and function

Types of dysbiosis



Related diseases in people



Gebrayel, P, et al. J Transl Med 20, 111 (2022)

Intestinal dysbiosis in inflammatory bowel disease

Nirmal Kaur,^{1,†} Chun-Chia Chen,^{2,†} Jay Luther¹ and John Y. Kao^{1,*}

Review

Gastrointestinal Microbiota and Type 1 Diabetes Mellitus: The State of Art

Marilena Durazzo *, Arianna Ferro and Gabriella Gruden

Review

Role of gut microbiota in type 2 diabetes pathophysiology

Manoj Gurung^{a,1}, Zhipeng Li^{a,1}, Hannah You^{a,1}, Richard Rodrigues^b, Donald B Jump^c, Andrey Morgun^{b,*}, Natalia Shulzhenko^{a,*}

Review: The Role of Intestinal Dysbiosis in Parkinson's Disease

Yiying Huang^{1†}, Jinchi Liao^{1†}, Xu Liu¹, Yunxiao Zhong¹, Xiaodong Cai^{2*} and Ling Long^{1*}

Circulation Research

REVIEW

Gut Microbiota and Cardiovascular Disease

Marco Witkowski, Taylor L. Weeks, Stanley L. Hazen

Related diseases in dogs & cats

Antibiotic therapy Exocrine pancreas insufficiency (EPI) Chronic inflammatory enteropathies

Food indiscretion/acute enteropathies

Obesity Diabetes mellitus CKD Atopic dermatitis

TOPIC HIGHLIGI

WJG 20th Anniversary Special Issues (17): Intestinal microbiota

Microbiota alterations in acute and chronic gastrointestinal inflammation of cats and dogs

Julia B Honneffer, Yasushi Minamoto, Jan S Suchodolski

Diabetic cats have decreased gut microbial diversity and a lack of butyrate producing bacteria

Ida Nordang Kieler¹, Melania Osto², Leoni Hugentobler², Lara Puetz³, M. Thomas P. Gilbert^{3,4}, Torben Hansen⁵, Oluf Pedersen⁵, Claudia E. Reusch⁶, Eric Zini^{6,7}, Thomas A. Lutz² & Charlotte Reinhard Bjørnvad¹

Microbiota-Related Changes in Unconjugated Fecal Bile Acids Are Associated With Naturally Occurring, Insulin-Dependent Diabetes Mellitus in Dogs

Albert E. Jergens^{1*}, Blake C. Guard², Alana Redfern¹, Giacomo Rossi³, Jonathan P. Mochel⁴, Rachel Pilla², Lawrance Chandra⁴, Yeon-Jung Seo⁴, Joerg M. Steiner², Jonathan Lidbury², Karin Allenspach¹ and Jan Suchodolski²



MDPI

Communication

Comparison of the Gut Microbiome between Atopic and Healthy Dogs—Preliminary Data

Ana Rostaher ^{1,*}, Yasser Morsy ²⁽⁰⁾, Claude Favrot ¹, Stefan Unterer ¹, Manuela Schnyder ³⁽⁰⁾, Michael Scharl ² and Nina Maria Fischer ¹

Diagnosing dysbiosis in practice

	Function	normal in Dogs	normal in Cats	Change in dysbiosis
aecalibacterium	anti-inflammatory, production of SCFA	3.4 - 8.0	3.8 - 8.4	\downarrow
Turicibacter	production of SCFA	4.6 – 8.1	4.4 - 9.0	\downarrow
Blautia	production of SCFA	9.5 – 11.0	not measured	\checkmark
usobacterium	production of SCFA	7.0 – 10.3	not measured	\checkmark
Bifidobacterium	production of SCFA	not measured	3.2 - 8.7	\downarrow
Bacteroides	production of SCFA	not measured	4.0 – 7.5	\checkmark
Clostridium hiranonis	conversion of primary to secondary bile acids	5.1 – 7.1	4.5 – 7.1	\downarrow
Streptococcus	overgrowth associated with dysbiosis	1.9 – 8.0	1.6 – 5.2	\uparrow
. coli	pro-inflammatory	0.9 – 8.0	1.4 – 7.0	↑

Validated in dogs and cats!

RESEARCH ARTICLE

A dysbiosis index to assess microbial changes in fecal samples of dogs with chronic inflammatory enteropathy

MK AlShawaqfeh^{1,2}, B Wajid^{1,3}, Y Minamoto¹, M Markel¹, JA Lidbury¹, JM Steiner¹, E Serpedin² and JS Suchodolski^{1,*}

A dysbiosis index to evaluate the fecal microbiota in healthy cats and

cats with chronic enteropathies

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(S)SAGE

Ims

Chi-Hsuan Sung¹, Sina Marsilio², Betty Chow^{3,4}, Kailee A Zornow⁶, Jennifer E Slovak⁶, Rachel Pilla¹, Jonathan A Lidbury¹, Jörg M Steiner¹, So Young Park¹, Min-Pyo Hong¹, Steve L Hill^{3,8} and Jan S Suchodolski¹





Fecal DI – in practice



Potential uses of fecal DI

Identifying sub-clinical dysbiosis in patients at risk

Identifying persistent antibiotic-induced dysbiosis

Based on severity and pattern (i.e. loss of *P. hiranonis*)

Increase likelihood of chronic enteropathy in patient with recurrent/acute on chronic GI signs (especially cats)

Predict long-term response to FMT

Screen for healthy fecal donors

Managing intestinal dysbiosis





Choice of strategy will depend on type/cause, duration and severity of dysbiosis

Multimodal versus sequential approach

Target (clinical remission versus cure) will depend on cause of dysbiosis

Treatment option	Mechanisms of action	Potential limitations
Diet	 Improved digestibility - less substrates for bacterial growth Reduced immunogenicity 	Palatability, cost
Prebiotics/fibers	Stimulates growth of beneficial bacteriaBinds toxic metabolites	Excess might cause clinical signs
Probiotics	Improve barrier functionImmunomodulation	Minimal effect on total microbiota composition
Antibiotics	 Reduction in total and mucosa-adherent bacteria Reduced toxic metabolites 	 Negative impact on microbiota diversity and beneficial species GI toxicity Limited effect in the long term Antimicrobial resistance
Fecal microbiota transplantation	 Modification of luminal microbiota and metabolites Reduction of enteropathogens 	 Minimal effect on mucosal adherent microbiota Limited duration of effect if underlying intestinal inflammation still present

Fecal microbiota transplantation (FMT)

Infusion of **fecal matter** from a healthy donor into the gastrointestinal tract of a patient (to introduce or re-establish a stable microbial community) in order to treat a dysbiosis-related disease

Super poo: the emerging science of stool transplants and designer gut bacteria







A recent FDA approval is likely only the start of a promising future for prescription poop.

* BY KATIE MACBRIDE DEC. 19, 2022

Major indication in people

Recurrent *Clostridiodes difficile infection* (CDI)

Acquired after antibiotic use + ingestion of environmental *CD* spores

Risk of relapse 20% Chronic long-lasting CDI (antibiotic refractory)

Annual burden > 80,000\$/patient

FMT therapeutic success rates > 90% Very high safety profile



Borody TJ et al, Nat Rev Gastroenterol Hepatol. 2011: 20;9(2):88-96

Milestone

FDA NEWS RELEASE

FDA Approves First Fecal Microbiota Product

Rebyota Approved for the Prevention of Recurrence of Clostridioides difficile Infection in Adults



FDA Approves First Orally Administered Fecal Microbiota Product for the Prevention of Recurrence of Clostridioides difficile Infection



Ongoing trials in people

Inflammatory bowel disease (IBD)

Irritable bowel syndrome (IBS)

Decolonisation from MDR organisms

Obesity

Metabolic syndrome

Neurological disorders

Autoimmune syndromes

Atopy and allergy

Liver disease and hepatic encephalopathy

Cancer

Chemo- and immunotherapy related GI signs

Review

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Fecal microbiota transplantation in gastrointestinal and extraintestinal disorders

Future

MICROBIOLOGY

Gianluca Ianiro*:¹, Jonathan P Segal², Benjamin H Mullish³, Mohammed N Quraishi^{4,5}, Serena Porcari¹, Ginevra Fabiani¹, Antonio Gasbarrini¹ & Giovanni Cammarota¹

Efficacy of FMT in dogs and cats

Infectious diarrhea (Parvovirus) - dogs

Acute uncomplicated diarrhoea – dogs

Antibiotic-induced dysbiosis and diarrhoea – dogs, cats

Chronic GI signs refractory to diet/fibre - dogs, cats

SRE and NRE as adjunctive therapy (multimodal therapy) – dogs, cats

Retrospective Under-powered Uncontrolled studies Clinical experience

Canine parvoviral enteritis

Pereira et al, 2018

66 puppies with CPVI

- 33 STD vs 33 STD + FMT
- Faster resolution of diarrhoea in FMT
- Shorter hospitalisation time
 - 3 vs 6 days

No statistical difference in survival rates

For FMT, 10 g of feces from a healthy dog diluted in10 mL of saline were administered rectally 6-12 hours post-admission



Pereira GQ, et al. J Vet Intern Med. 2018 Mar;32(2):707-711

Canine chronic enteropathy (CE)

Toresson et al, 2022

Retrospective study 41 dogs with SRE

Treated via – retention enema Median 3 FMTs 14 days apart

72% of dogs showed satisfactory clinical response

Degree of dysbiosis (DI) associated with response to FMT



Toresson L, et al. Veterinary Sciences. 2023; 10(4):271

Canine food refractory CE

- Vecchiato C et al (ECVIM Congress 2023) in press
- 20 dogs with idiopathic chronic GI signs
- No response to 2-weeks trial with hydrolysed or single protein home cooked diet
- 8 x 1 FMT 12 x 2 FMTs
- **Clinical response in 20/20**
- FCS from 4 (1-7) to 2 (1-5) at 60 days
- CIBDAI from 5 (1-9) to 1 (0-3) at 90 days
- DI from 0.1 (range -5.6 to 3.8) 2.1 (range -5.7 to 4.7)





First Case Report of Fecal Microbiota Transplantation in a Cat in Israel

Furmanski, S.^{1,*} and Mor, T.²

FMT (enema) as a last therapeutic option for cat before euthanasia

Immediate improvement in fecal texture, odor and color

Second FMT was performed 5 weeks later for relapse Over a 3-month period gradual return to normal stools

11 months after 2nd FMT cat in remission



Furmanski S. and Mor T. Israel Journal of Veterinary Medicine Vol. 72 (3) 2017; 35-41



Feline antibiotic induced dysbiosis

Jamie Hui et al, ACVIM abstract 2022

Healthy cats treated for 10 days with amoxyclav

FMT via enema (**blue**) or placebo (**black**) after last day of antibiotic



Clinical guidelines

Companion Animal FMT Consortium

FMT Guidelines for Clinical Practice

Donor Screening & Selection

FMT Preparations

FMT Dosing & Clinical Applications

ARTICLE IN PRESS

Advances in Small Animal Care = (2024) =-= ADVANCES IN SMALL ANIMAL CARE

Clinical Guidelines for Fecal Microbiota Transplantation in Companion Animals

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Preparation

Fresh (within 2-6 hours from voiding)

Fresh-frozen (processed as soon as possible after voiding

- can be kept refrigerated at 4°C for up to 2 days before)

Addition of cryopreservant before freezing (glycerol 1:10)

Processing can be done in aerobic environment



My method

2.5-5 gr feces /kg b.w. of recipient

Blend in 1:1 or 1:2 with 0.9% NaCl solution by hand in a zip bag

Filtered on fine kitchen sieve

Freeze after adding glycerol 1:10 for later use

Administer after thawing with 20-60 cc syringe through wide bore soft rectal catheter (12 Fr)

With or without sedation (usually needed in cats)

No food and no walk for 4-6 hours (dogs) Keep in hospital for 4-6 hours (cats)





Repeat 1-2 weeks apart



Some real life case examples



Meet Chloe

5 yrs old FN Maltese cross

Refractory/non-responsive chronic enteropathy

Life long intermittent GI signs Continuously over last 2 months

Previously: Fiber enriched diet & novel protein diet trials Metronidazole

Advised endoscopy for GI biopsies and glucocorticoids trial



At the time of presentation

Daily vomiting (mostly bilious) Liquid feces with frank blood and mucus (PFS 5-7/7) & tenesmus

Nocturnal swallowing fits Selective appetite Fed on chicken or pork based home cooked diet

Stable weight, BCS 5/9





Diagnostic and clinical staging

Severe clinical disease activity (CIBDAI score 9)

Fecal flotation - negative

CBC - mild eosinophilia **Serum biochem** - Mild ALT and DGGR lipase activities increase cTLI and basal cortisol ruled out EPI and HypoA

Serum cobalamin < 150 ng/L

Abdominal ultrasound: indicative of non-specific mild chronic enteropathy

Initial treatment strategy

Awaiting - fecal DI

Eliminate all extras from daily diet

Hydrolysed soy diet

Multistrain high dose probiotic

Oral cobalamin 0.5 mg PO q 24 hours

Two weeks follow-up

Partial response

CIBDAI 3

Purina fecal score 3/7

Occasional bilious vomiting + colitis





Fecal DI



Post FMT follow-up

Complete remission

CIBDAI 0

Purina fecal score 2/7

Weight gain

No more vomiting episodes





Follow-up DIs



Skoll

3 yrs old ME Alaskan Malamute

Referred for refractory chronic mixed bowel diarrhoea – 3 months duration

Failed hydrolysed (soy) + single protein (horse) home cooked diets trials

Gastroduodenal-colonoscopy + H&E: mild to moderate lymphoplasmacytic enterocolitis

Minimal response to glucocorticoids + probiotics (still receiving)



At the time of presentation

Severe mixed bowel diarrhoea (FS 7/7) Hematochezia + mucus Increased number of defecation

Dysorexia Rare vomiting episodes

Weight and muscle loss BCS 4/9

Polyuria, polydipsia, poor exercise tolerance







Staging

Severe clinical disease activity (CIBDAI score 11)

CBC: within normal limits

Biochemistry: moderate ALT and ALP increase, marked DGGR lipase increase, mild CRP increase

cTLI: 33 ug/L Serum cobalamin: 588 ng/L

Abdominal ultrasonography: small sized adrenal glands, vacuolar (steroid) hepatopathy, mild chronic enteropathy

Baseline DI



Initial treatment plan

Highly digestible low residue fiber enriched diet

Taper down glucocorticoids

Continue high dose multistrain probiotic



Follow-up

Off glucocorticoids

CIBDAI 5 (mild disease activity)

FS 5/7 Occasional hematochezia and mucus Frequency of defecation reduced

Improved appetite and exercise tolerance Weight gain

PU/PD only mild





Follow-up DIs



After FMT

Off glucocorticoids

CIBDAI 0 FS 2-3/7 No other clinical signs

Excellent appetite 2 kg body weight increase PU/PD resolved





Follow-up DIs



2 yrs old MN DSH

Chronic small intestinal diarrhea, 2-3 times a day No identifiable cause

Refractory to fiber enriched-, highly digestible, hydrolyzed diets and probiotics

Responsive to amoxicillin clavulanate

Gradual return to liquid stools when discontinued Lost responsiveness with time

Lupin

Fecal DI – 1 month off antibiotics

DI	2.2	< 0
Clostridium hiranonis	0.8	4.5-7.1
Bacteroides	5.9	4.0-7.5
Bifidobacterium	6.2	3.2-8.7
Turicibacter	4.8	4.4-9.0
Faecalibacterium	3.9	3.8-8.4
Streptococcus	2.8	1.6-5.2
E. coli	3.9	1.4-7.0

https://vetmed.tamu.edu/gilab/

Pre- and post-FMT Dysbiosis index

DI	2.2	0.8	< 0
Clostridium hiranonis	0.8	4,4	4.5-7.1
Bacteroides	5.9	5.3	4.0-7.5
Bifidobacterium	6.2	7.1	3.2-8.7
Turicibacter	4.8	6.4	4.4-9.0
Faecalibacterium	3.9	4.0	3.8-8.4
Streptococcus	2.8	2.2	1.6-5.2
E. coli	3.9	5.4	1.4-7.0

1 + 4 weeks post FMT

Take home messages

Treatment strategies for intestinal dysbiosis should depend on the underlying cause and will often require a multimodal approach

Cure is possible in mild cases (provided the eliciting cause is eliminated) or in antibiotic-induced dysbiosis (as sole treatment)

Clinical remission is a reasonable target in CE-associated dysbiosis (multimodal strategy)

Based on current available evidence, a dietary trial should be attempted first in dogs and cats with chronic GI signs and should always be part of multimodal treatment strategies

Take home messages

Besides inducing antimicrobial resistance, antibiotics are detrimental to gut and microbiome health

They should not be routinely used for the diagnostic approach and treatment of GI signs

FMT appears to be efficacious especially in young dogs and cats with mild dysbiosis or antibiotic-induced dysbiosis and associated GI signs

The use of FMT in CE appears promising for clinical remission, especially in the context of a multimodal approach

+ + + + + + CREATING CLARITY

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Thank you very much for the attention!

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Questions?

